

Fungal Pathogens on Some Lepidopteran Forest Pests in Bulgaria

S. Draganova¹, D. Takov², D. Pilarska^{2,5}, D. Doychev³, P. Mirchev⁴, G. Georgiev⁴

¹Institute of Soil Science, Agrotrecheologies and Plant Protection, 7 Shosse Bankya Str., 1080 Sofia, Bulgaria; E-mail: sdraganova19@gmail.com

²Institute of Biodiversity and Ecosystem Research, Bulgarian Academy of Sciences, 2 Gagarin Str., 1113 Sofia, Bulgaria; E-mails: dpilarska@yahoo.com; dtakov@yahoo.com

³University of Forestry, 10, Kl. Ohridski Blvd., Sofia 1756, Bulgaria; E-mail: doychev@abv.bg

⁴Forest Research Institute, Bulgarian Academy of Science, 132 St. Kliment Ohridski Blvd., 1756 Sofia; Bulgaria;

⁵Czech University of Life Sciences, 129 Kamýcká str, 165 21 Prague 6, Suchdol, Czech Republic; E-mail: dpilarska@yahoo.com

Abstract: Fungal entomopathogens found in natural populations of 7 lepidopteran forest pests were isolated and identified. *Beauveria bassiana* was established as a dominating species followed by *Fusarium* sp. and *Aspergillus* sp. The pine processionary moth *Thaumetopoea pityocampa* was the most affected by *B. bassiana* – 66.7% of the analysed dead larvae of the pest. The occurrence of *B. bassiana* in *Malacosoma neustria*, *Tortrix viridana* and *Melitaea didyma* is the first record in Bulgaria.

Relative susceptibility of first-second instar *Lymantria dispar* larvae was estimated to three *B. bassiana* and one *Metarhizium anisopliae* isolate and susceptibility of third instars – to seven *B. bassiana* and one *M. anisopliae* isolate. The bioassays showed that *L. dispar* larvae were tolerant to mycoses caused by the tested *B. bassiana* isolates and *M. anisopliae*. *M. anisopliae* isolate caused mycosis to first-second instar larvae with the highest lethal effect $56.67\% \pm 4.84$ and *B. bassiana* isolates 561Bb and 575Bb to third instar larvae – $47.78\% \pm 4.06$ and $34.44\% \pm 3.28$, respectively, in 17 days period.

Key words: lepidopteran forest pests, entomopathogenic fungi, *Lymantria dispar*, *Beauveria bassiana*, *Metarhizium anisopliae*

Introduction

Lepidopteran species such as pine processionary moth *Thaumetopoea pityocampa* (Den. & Schiff.), the gypsy moth *Lymantria dispar* (L.), green oak leafroller *Tortrix viridana* L., *Malacosoma neustria* (L.), *Euproctis chrysorrhoea* (L.) and Geometridae species *Operophtera brumata* (L.) and *Erannis defoliaria* (Cl.) are among the most harmful pests in coniferous and deciduous forests registered in Bulgaria (MIRCHEV, TSANKOV 2000). Their mass outbreaks could cause serious damages even to defoliate deciduous trees.

The existent IPM of lepidopteran forest pests include aerial spray treatments using several preparations of *Bacillus thuringiensis* var. *kurstaki* or

chemical pesticides like Dimilin. Efforts to improve the existent environment friendly strategies were the reason to study fungal pathogens – their biodiversity (PAPIEROK *et al.* 1984, SOLTER *et al.* 2000, 2002, JANKEVICA 2004, MIRCHEV 2004, PILARSKA *et al.* 2001, 2006, 2010; POLOVINKO *et al.* 2010; SEVIM *et al.* 2010), pathogenicity (ER *et al.* 2007; POLOVINKO *et al.* 2010; SEVIM *et al.* 2010) and to conduct introduction of the entomophthorous fungus *Entomophaga mai-maiga* Humber, Shimazu & Soper (PILARSKA *et al.* 2000, 2007, 2010).

The aim of the study was to establish naturally occurring fungal pathogens on some lepidopteran forest pests in Bulgaria and to estimate the relative

susceptibility of first-second and third instar *L. dispar* larvae to 7 isolates of *Beauveria bassiana* (Bals. – Criv.) Vuillemin and one isolate of *Metarhizium anisopliae* (Metsch.) Sorokin.

Materials and Methods

Natural occurrence

The occurrence of fungal entomopathogens in natural populations of lepidopteran forest pests was studied in 2 year period from April 2009 to June 2011. A total of 563 individuals of insects (larvae, pupae, adults) belonging to 7 lepidopteran species were collected from 12 forest stands from different regions in Bulgaria (Table 1).

The found dead insects were collected in individual containers for laboratory analyses. The collected alive larvae were reared on natural diet (leaves of apple, plum or oak trees) in the laboratory till their death or imagination. All dead insects were analysed microscopically. Parts of cadavers of insects with symptoms of mycosis after surface sterilization were placed in a humid chamber at temperature of 25 ± 1 °C for sporulation of fungal pathogens. Conidia from sporulated fungi were transferred to SDAY (Sabouraud dextrose agar with yeast extract) in glass Petri dishes and cultured for 15 days at 25 ± 2 °C at a photoperiod of L12:D12. The obtained pure cultures of the isolates after subculturing on slopes of the same media in tubes were kept in a refrigerator at temperature of 4 ± 2 °C. Conidia from pure cultures of isolates and small parts of cadavers were used to prepare smears stained by methylen-blue as well as slide preparations with lactophenol and aniline blue closed by nail vanish (after HUMBER 1997). Conidia and conidiogenous cells were observed using a transmission interference microscope BX60 DIC Olympus equipped with digital camera and Cell B image capture software with calibrated Carl Zeiss micrometer. The fungal pathogens were identified according to their macro- and micro-morphological characteristics according to SAMSON *et al.* (1988) and HUMBER (1997).

Bioassays with fungal isolates and larvae of *L. dispar*

Seven isolates of *B. bassiana* obtained from different lepidopteran and coleopteran forest insects and one isolate of *M. anisopliae* from a dead adult of the grey corn weevil were used in bioassays

(Table 2). They were cultured for 15 days on slopes of SDAY in tubes under the conditions mentioned above and obtained conidia were washed down with sterilized water containing 0.05% Tween 80. The concentrations of conidia were determined by enumeration of conidia in Bürker chamber after serial dilution of aqueous suspensions. Suspensions applied in bioassays were prepared at a concentration of 1×10^8 conidia/ml.

First-second and third instar *L. dispar* larvae used to estimate the relative susceptibility to fungal isolates were from a laboratory population reared on natural diet (plum-tree leaves) at temperature of 22 ± 1 °C and $70 \pm 1\%$ RH. Insects were obtained after hatching of gypsy moth eggs provided by the USDA-APHIS-PPQ Laboratory, Buzzards Bay, MA, USA. The insects were treated by a surface contact with 1 ml of conidial suspensions (1×10^8 conidia/ml) placed on filter paper discs (90 mm in diameter) in glass Petri dishes (DRAGANOVA, STANEVA 1990). The larvae in control variants were treated with 1 ml of water containing 0.05% Tween 80 instead of conidial suspension. The filter paper disks were removed 24 h after the treatments and insects were fed on plum-tree leaves. The experiments were conducted under laboratory conditions at temperature of 22 ± 1 °C and $70 \pm 1\%$ RH in three replicates with 30 first-second or third instar *L. dispar* larvae per a replicate. Insect mortality was checked daily for 17 days after contamination. Dead insects in all treatments were removed daily and were placed on moist filter paper in new Petri dishes for a fungal pathogen exhibition. The results of the bioassays were evaluated as percentages of cumulative daily mortality due to mycoses, corrected with mortality in the control treatment following the Schneider-Orelli's formula (PÜNTENER 1981). Differences among lethal effects of mycoses to *L. dispar* larvae in treatments with conidial suspensions of fungal isolates were estimated by t-test for independent samples applying Descriptive statistics. Values of $p < 0.05$ were considered significant. Statistical analyses were performed using the software STATISTICA^R version 6.0 of Stat Soft Inc.

Results and Discussion

Natural occurrence

The analyses of 145 dead individuals from 7 lepidopteran species showed that 89 of them were with symptoms of mycoses (Table 3). Fungal patho-

Table 1. Lepidopteran forest pests.

Species, family	Locality
<i>Thaumetopoea pityocampa</i> (Den. et Schiff.), Notodontidae	Banya (Karlovo district)
<i>Lymantria dispar</i> L., Lymantriidae	Pobit kamak (Sredna Gora Mountain) Konush (Eastern Rhodopes) Perperek (Eastern Rhodopes) Gnyazdovo (Eastern Rhodopes) Krumovgrad (Eastern Rhodopes) Nanovitza (Eastern Rhodopes) Zvezdel (Eastern Rhodopes) Kamenetz (Eastern Rhodopes)
<i>Aporia crataegi</i> L., Pieridae	Kamenetz (Eastern Rhodopes)
<i>Malacosoma neustria</i> L., Lasiocampidae	Gnyazdovo (Eastern Rhodopes) Kamenetz (Eastern Rhodopes)
<i>Erannis defoliaria</i> Clerck, Geometridae	Gnyazdovo (Eastern Rhodopes) Perperek (Eastern Rhodopes)
<i>Melitaea didyma</i> Esper, Nymphalidae	Krumovgrad (Eastern Rhodopes)
<i>Tortrix viridana</i> L., Tortricidae	Plakovo (Central Balkan Range) Pobit kamak (Sredna Gora Mountain) Kardjali (Eastern Rhodopes) Perperek (Eastern Rhodopes) Gnyazdovo (Eastern Rhodopes) Komuniga (Eastern Rhodopes) Krumovgrad (Eastern Rhodopes) Zvezdel (Eastern Rhodopes)

Table 2. Origin of the isolates of entomopathogenic fungi applied in bioassay with *Lymantria dispar* larvae (L₁₋₂ and L₃)

Isolate, entomopathogenic fungus	Origin (initial insect host)		Year of isolation
	Species	Stage	
560Bb, <i>B. bassiana</i>	<i>Thaumetopoea pityocampa</i> (Den. et Schiff.) (Lepidoptera: Notodontidae)	larva	2009
561Bb, <i>B. bassiana</i>	<i>Stenomax aeneus</i> (Scop.) (Coleoptera: Tenebrionidae)	imago	2009
575Bb, <i>B. bassiana</i>	<i>Lymantria dispar</i> L. (Lepidoptera: Lymantriidae)	larva	2009
588Bb, <i>B. bassiana</i>	<i>Tortrix viridana</i> L. (Lepidoptera: Tortricidae)	larva	2010
593Bb, <i>B. bassiana</i>	<i>Operophtera brumata</i> L. (Lepidoptera: Geometridae)	larva	2010
596Bb, <i>B. bassiana</i>	<i>Thaumetopoea pityocampa</i> (Den. et Schiff.) (Lepidoptera: Notodontidae)	larva	2010
601Bb, <i>B. bassiana</i>	<i>Thaumetopoea pityocampa</i> (Den. et Schiff.) (Lepidoptera: Notodontidae)	larva	2010
619Ma, <i>M. anisopliae</i>	<i>Tanymecus dilaticollis</i> Gyll. (Coleoptera: Curculionidae)	imago	2010

gens were identified as *Beauveria bassiana* (Bals. – Criv.) Vuillemin (Sordariomycetes: Hypocreales, Cordycipitaceae), *Fusarium* sp. (Sordariomycetes, Hypocreales, Nectriaceae) and *Aspergillus* sp. (Eurotiomycetes, Eurotiales, Trichocomaceae). Fifty seven *B. bassiana* isolates were obtained in pure cultures on SDAY. The established results revealed that *B. bassi-*

Table 3. Pathogens established in natural populations of lepidopteran forest pests

Lepidopteran host			Pathogen (number of infected individuals)
Species, family	Total number of investigated individuals	Number of dead individuals	
<i>Thaumetopoea pityocampa</i> (Den. et Schiff.), Notodontidae	39 L 31 P	33 L 11 P	<i>B. bassiana</i> (22 L) <i>Fusarium</i> sp. (5 L), <i>Aspergillus</i> sp. (6 L, 7 P) Other (4 P)
<i>Lymantria dispar</i> (L.), Lymantriidae	59 L 8 P 2 i	33 L 3 P 1 i	<i>B. bassiana</i> (4 L, 1 P) <i>Aspergillus</i> sp. (2 L, 2 P) Other (27 L, 1 i)
<i>Malacosoma neustria</i> (L.), Lasiocampidae	131L	29L	<i>B. bassiana</i> (9 L) <i>Fusarium</i> sp. (2 L) Other (18 L)
<i>Tortrix viridana</i> L., Tortricidae	238 L 34 P 3 i	16 L 5 P 2 i	<i>B. bassiana</i> (10 L, 2 P, 2 i) <i>Fusarium</i> sp. (1 L) <i>Aspergillus</i> sp. (5 L) Other (3 P)
<i>Erannis defoliaria</i> (Cl.), Geometridae	16 L	10 L	<i>B. bassiana</i> (6 L) <i>Aspergillus niger</i> (2 L) Other (2 L)
<i>Aporia crataegi</i> (L.), Pieridae	1 L	1 L	Other (1 L)
<i>Melitaea didyma</i> (Esp.), Nymphalidae	1 i	1 i	<i>B. bassiana</i> (1 i)

*L – larva, P – pupa, i – imago

**Other – reason of death different from mycosis

ana was the fungal pathogen that occurred in natural populations of 6 from 7 species of the observed lepidopteran forest pests.

The larvae of the pine processionary moth were the most affected by the fungus – 66.7% of the analysed dead larvae. Twenty two *B. bassiana* isolates were obtained in pure cultures from them. The reason of such high extensity observed was that *B. bassiana* as soil-dwelling fungus can easily come into contact with the premature stages of *T. pityocampa* during their staying in the soil and infect them. Fungi are active pathogens which attack their hosts percutaneously. They produce extracellular enzymes lysing the host cuticula and penetrate through the arthropod integument with support of mechanical pressure of the growing tubes of the germinating conidia (ST. LEGER *et al.* 1986, DRAGANOVA 1988, GUPTA *et al.* 1992; ST. LEGER 1995).

Our investigations established that *B. bassiana* occurred as a causal agent of mycoses in natural populations of *L. dispar* (7.2% of all collected and 13.5% of dead individuals), *M. neustria* (6.9% of

all and 31% of dead individuals), *T. viridana* (5.1% of all and 60.9% of dead individuals), *E. defoliaria* (37.5% of all observed and 60% of dead individuals). Dead by mycoses were mainly larvae, rarely pupae. Few adults of some lepidopteran hosts (*T. viridana* and *M. didyma*) were found to be affected by the fungal pathogen as well (Table 3). This is the first record of *B. bassiana* in *M. neustria*, *T. viridana* and *M. didyma* in Bulgaria.

PANAJOTOV *et al.* (1960) reported that the fungi *B. bassiana*, *B. glubulifera* Pic. and *Spicaria farinosa* Vuill. (now *Isaria farinosa* (Holmsk.) Fries) were found on *L. dispar* larvae in Bulgaria. HAJEK *et al.* (1997) found out that *Paecilomyces farinosus* (Holmsk. ex Gray) Brown & Smith) (now *I. farinosa*) was the most often occurring hyphomycete species in natural populations of gypsy moth although in low infection level (4.6% – 12.2%) and rarely *B. bassiana*. According to GOETTEL *et al.* (1990) and BUTT *et al.* (2001) *B. bassiana* is an entomopathogenic fungus with a large host range which includes about 700 arthropod species.

POLOVINKO *et al.* (2010) reported that *B. bassiana* was a dominating species comprising on average 68% of the total number of isolates from Lepidoptera and Coleoptera in West Siberia, Primorsky Krai, and Kyrgyzstan. During observation period the mortality rate of the insects died of *B. bassiana* was registered mostly on enzootic level although in some cases the local nidi of mass death of the hosts were noted. *Isaria farinosa* (Holmsk.) Fries was the entomopathogenic fungus found primarily on Lepidoptera – the Macroheterocera group. According to POLOVINKO *et al.* (2010) *Isaria farinosa* was also likely to have a significant epizootic potential. It was revealed that in some places in the region of Novosibirsk the mentioned fungus was present in soil in great amounts.

Fungal entomopathogens found to cause mycoses in natural populations of *T. pityocampa* were studied by SEVIM *et al.* (2010). Based on their morphological and molecular characteristics four isolates were identified by the authors as *Beauveria bassiana* cf. Clade C, and one isolate was identified as *B. bassiana*.

The occurrence of *B. bassiana* in natural populations of *T. pityocampa* and *L. dispar* in Bulgaria and study of some properties of the obtained fungal isolates was reported by DRAGANOVA *et al.* (2011).

Another representative of genus *Beauveria* – *B. brongniartii* (Saccardo) Petch was announced by JANKEVICA (2004) to infect European tent caterpillar *M. neustria* in a study on the occurrence of entomopathogenic fungi and their host range.

The fungal species from genera *Fusarium* and *Aspergillus* found by us on insect cadavers (Table 3) are considered as weak pathogens and not to be the

reason for the death of insects. The infected insects weakened by starving or injuring and rarely appear in natural host populations. According to EVLAKHOVA (1974) species of genus *Aspergillus* were not typical entomopathogens and even according to Humber (1998) they were contaminants growing on cadavers after insect death. Opposite to them BENNETT (2010) considered that *A. flavus* Link was capable of causing serious disease to plants, insects and vertebrates taking into account that it produced aflatoxin and was a pan-kingdom pathogen.

MIRCHEV (2004) found that 24.5% of mortality of *L. dispar* pupae was due to diseases caused by *Scopulariopsis breviculalis* (Sacc.) Bain, *A. flavus*, *Penicillium frequentans* Westl., *Mucor mucedo* Fres., *M. globosus* Fischer, *Aspergillus* sp. and *Fusarium* sp.

The analyses of the found dead or diseased lepidopteran forest forest pests showed that except mycoses there were another reasons for the decreased population density (Table 3). We marked them as „others“ and they presented 38.6% of all cases of insect death. As a whole it could be mentioned that they were with different causal agents – virus, bacteria, parasitoids or predators, but they will be object of our future study.

Bioassays with *L. dispar* larvae

The results obtained in bioassays revealed that first-second instar *L. dispar* larvae were tolerant to mycoses caused by the examined *B. bassiana* isolates 588Bb, 593Bb and 596Bb. At the 17th day after the treatments, values of cumulative daily mortality due to mycoses reached hardly to 14.44% \pm 1.28, 15.56% \pm 1.15 and 14.44% \pm 1.26, respectively

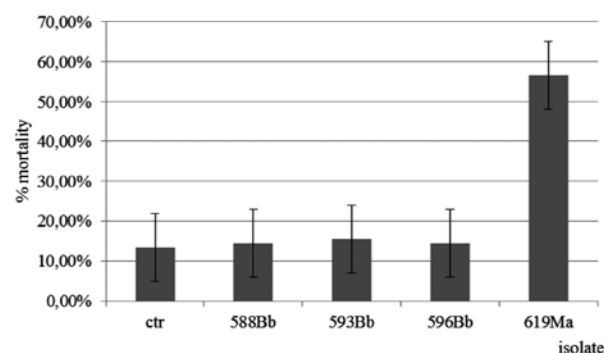


Fig. 1. Lethal effect of mycoses caused by 3 *Beauveria bassiana* isolates (588Bb, 593Bb, 596Bb) and one isolate of *Metarhizium anisopliae* (619Ma) to first-second instar *Lymantria dispar* larvae established at the 17th day after the treatment with conidial suspensions (1×10^8 conidia/ml)

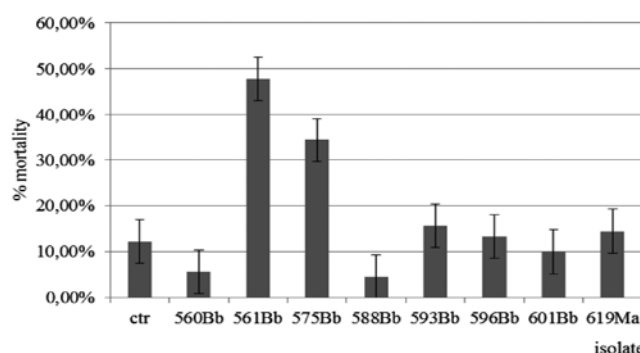


Fig. 2. Lethal effect of mycoses caused by isolates of *Beauveria bassiana* (560Bb, 561Bb, 575Bb, 588Bb, 593Bb, 596Bb, 601Bb) and *Metarhizium anisopliae* (619Ma) to third instar *Lymantria dispar* larvae established at the 17th day after the treatment with conidial suspensions (1×10^8 conidia/ml)

Table 4. Differences between mean lethal effects of mycoses to first-second instar *L. dispar* larvae in treatments with conidial suspensions (1×10^8 conidia/ml) of 4 *B. bassiana* isolates and one isolate of *M. anisopliae*

Variants compared	Mean lethal effect Group 1	Mean lethal effect Group 2	t	df	p	t separ. var.est.	df	p 2-sided	Valid N Group 1	Valid N Group 2	Std.Dv. Group 1	Std.Dv. Group 2	F-ratio variances	p variances
588Bb_L1 vs. CTR_L1	0,0732	0,0510	1,1092	32	0,2756	1,1092	30,9365	0,2759	17	17	0,0527	0,0636	1,4552	0,4614
593Bb_L1 vs. CTR_L1	0,0837	0,0510	1,7005	32	0,0987	1,7005	29,5648	0,0994	17	17	0,0473	0,0636	1,8050	0,2482
596Bb_L1 vs. CTR_L1	0,0876	0,0510	1,8361	32	0,0757	1,8361	30,8062	0,0760	17	17	0,0521	0,0636	1,4902	0,4337
619Ma_L1 vs. CTR_L1	0,3693*	0,0510*	6,2638*	32*	0,0000*	6,2638*	19,2124*	0,0000*	17*	17*	0,1996*	0,0636*	9,8600*	0,0000*
593Bb_L1 vs. 588Bb_L1	0,0837	0,0732	0,6095	32	0,5465	0,6095	31,6358	0,5465	17	17	0,0473	0,0527	1,2404	0,6717
596Bb_L1 vs. 588Bb_L1	0,0876	0,0732	0,8003	32	0,4294	0,8003	31,9955	0,4294	17	17	0,0521	0,0527	1,0240	0,9627
596Bb_L1 vs. 593Bb_L1	0,0876	0,0837	0,2292	32	0,8202	0,2292	31,7106	0,8202	17	17	0,0521	0,0473	1,2113	0,7061
619Ma_L1 vs. 588Bb_L1	0,3693*	0,0732*	5,9123*	32*	0,0000*	5,9123*	18,2194*	0,0000*	17*	17*	0,1996*	0,0527*	14,3484*	0,0000*
619Ma_L1 vs. 593Bb_L1	0,3693*	0,0837*	5,7396*	32*	0,0000*	5,7396*	17,7923*	0,0000*	17*	17*	0,1996*	0,0473*	17,7976*	0,0000*
619Ma_L1 vs. 596Bb_L1	0,3693*	0,0876*	5,6294*	32*	0,0000*	5,6294*	18,1678*	0,0000*	17*	17*	0,1996*	0,0521*	14,6935*	0,0000*

* difference between variants is significant

(Fig. 1). Larvae were relatively more susceptible to mycosis caused by the *M. anisopliae* isolate 619Ma. Calculated values of larval mortality established in the treatments with 619Ma were $56.67\% \pm 4.84$.

Differences between mean lethal effects of mycoses to first-second instar *L. dispar* larvae in treatments with conidial suspensions (1×10^8 conidia/ml) of fungal isolates were estimated by t-test for independent samples (Table 4). Significant difference was proved at $p\text{-level} < 0.05$ only between variant treated with conidial suspensions of 619Ma, control variant and treatments with *B. bassiana* isolates. The examined three *B. bassiana* isolates were not significantly different among each other by their lethal effect to first-second instar *L. dispar* larvae, and they were not different from the control variant as well.

The results of relative susceptibility estimation showed that the third instar *L. dispar* larvae were more tolerant to mycoses caused by the examined *B. bassiana* and *M. anisopliae* isolates than younger instars (Fig. 2). The established values of cumulative daily mortality due to mycoses were very low and only treatments with 561Bb and 575Bb caused mycoses with lethal effect $47.78\% \pm 4.06$ and $34.44\% \pm 3.28$, respectively.

Nevertheless the isolate 575Bb was obtained from dead *L. dispar* larva, it was with low virulence to larvae of its initial host. It seems that *L. dispar* is tolerant to mycoses caused by Hyphomycetes and especially by *B. bassiana* and *M. anisopliae* as reports about natural occurrence of these pathogens are few.

According to POLOVINKO *et al.* (2010), *L. dispar* larvae were less sensitive to the isolates of *B. bassiana*, which was confirmed by slow progress of the mycosis. The caterpillars of *L. dispar* died in 13–18 days after the treatment with selected 11 West Siberian isolates of the fungus *B. bassiana*. The authors considered that the isolates from a certain host were not necessarily more virulent for the same insect species.

Differences between mean lethal effects of mycoses to third instar *L. dispar* larvae in opposite to first-second instars were significantly proved at $p\text{-level} < 0.05$ in most of the cases.

Conclusions

The presented study reveals the distribution of hyphomycete entomopathogens in natural populations of 7 lepidopteran species in Bulgarian forests. *Beauveria bassiana* was established as a dominating species affecting individuals from larval, pupal and imaginal stages. The occurrence of *B. bassiana* in *Malacosoma neustria*, *Tortrix viridana* and *Melitaea didyma* is the first record in Bulgaria.

The bioassays showed that *L. dispar* larvae were tolerant to mycoses caused by the tested *B. bassiana* isolates and *M. anisopliae*. Future investigations should be directed to search for more virulent fungal isolates and perhaps new fungal pathogens as well.

Acknowledgements: We are especially indebted to the National Science Fund of Bulgaria, Project DO-02-251/2008, and to Prof. Leellen Solter for providing gypsy moth egg masses.

References

- BENNETT J. W. 2010 An Overview of the Genus *Aspergillus*. – In: M. MACHIDA and K. GOMI (Eds.): *Aspergillus – Molecular Biology and Genomics*, 1-17.
- BUTT T. M., C. JACKSON, N. MAGAN (Eds). 2001. *Fungi As Biocontrol Agents: Progress, Problems and Potential*, CABI Publishing, 390 p.
- DRAGANOVA S., D. PILARSKA, D. TAKOV, D. DOYCHEV. 2011. Utilization of carbohydrates by *Beauveria bassiana* isolates obtained from forest pests. – *Journal of Plant Protection Research*, **51** (3): 339-344.
- DRAGANOVA S., E. STANEVA 1990. Methods for screening strains of entomopathogenic fungi of *Beauveria* Vuill. genus by their virulence. – *Comptes rendus de l'Academie bulgare des Sciences*, **43** (8): 93-95.
- ER M., H. TUNAZ, A. GOKCE 2007. Pathogenicity of entomopathogenic fungi to *Thaumetopoea pityocampa* (Schiff.) (Lepidoptera: Thaumetopoeidae) larvae in laboratory conditions. – *Journal of Pest Science*, **80**: 235-239.
- EVLAKHOVA A. A. 1974. Entomogenous fungi. Classification, biology, practical significance. Nauka Press, Leningrad, 260 pp.
- GOETTEL M. S., T. J. POPRAWSKI, J. D. VANDENBERG, Z. LI, D. W. ROBERTS 1990. Safety to nontarget invertebrates of fungal biocontrol agents. – In: Laird M., L. A. Lacey and E. W. Davidson (Eds.): *Safety of microbial insecticides*, Boca Raton, FL: CRC Press, 209-232.
- GUPTA S. C., T. D. LEATHERS, G. N. EL-SAYED, C. M. IGNOFFO 1992. Insect cuticle-degrading enzymes from the entomopathogenic fungus *Beauveria bassiana*. – *Experimental Mycology*, **16**: 132-137.
- HAJEK A., J. ELKINTON, R. HUMBER 1997. Entomopathogenic Hyphomycetes associated with gypsy moth larvae. – *Mycologia*, **89** (6): 825-829.
- HUMBER R. 1997. Fungi: identification. – In: Lacey L. A. (Ed.): *Manual of techniques in insect pathology*, Academic Press, San Diego, USA, pp. 153-163.
- JANKEVICA L. 2004. Ecological associations between

- entomopathogenic fungi and pest insects in Latvia. – *Latvijas entomologs*, **41**: 60-65.
- MIRCHEV P. 2004. Longevity of *Lymantria dispar* L. at the pupa stage in low species population number. – *Forest Science*, **3**: 77-85.
- MIRCHEV P., G. TSANKOV 2000. Factors influencing on changes in the distribution and economic importance of the pine processionary moth (*Thaumetopoea pityocampa* Den. et Schiff.) in Bulgaria. – *Forest Science*, **2/3**: 15-24.
- PANAJOTOV P., B. ZASHEV, R. GRIGOROVA, G. TSANKOV 1960. Entomopathogenic fungi on caterpillars of *Lymantria dispar* L. in Bulgaria. – *Bulletin de l'Institut central des forests*, **6**: 205-208.
- PAPIEROK B., B. TORRES, M. ARNAULT 1984. Contribution a l'etude de la specificite parasitaire du champignon entomopathogene *Zoophthora radicans* (Zygomycetes, Entomophthorales). – *BioControl*, **29** (1): 109-119.
- PILARSKA D., M. MCMANUS, A. HAJEK, F. HERARD, F. VEGA, P. PILARSKI, G. MARKOVA 2000. Introduction of the entomopathogenic fungus *Entomophaga maimaiga* Hum., Shim. & Sop. (Zygomycetes: Entomophthorales) to a *Lymantria dispar* (L.) (Lepidoptera: Lymantriidae) population in Bulgaria. – *Journal of Pest Science (Anzeiger fur Schadlingskunde)*, **73** (5): 125-126.
- PILARSKA D., M. MCMANUS, P. PILARSKI, G. GEORGIEV, P. MIRCHEV, A. LINDE 2006. Monitoring the establishment and prevalence of the fungal entomopathogen *Entomophaga maimaiga* in two *Lymantria dispar* L. populations in Bulgaria. – *Journal of Pest Science*, **79** (2): 63-67.
- PILARSKA D., G. GEORGIEV, M. MCMANUS, P. MIRCHEV, P. PILARSKI, A. LINDE 2007. *Entomophaga maimaiga* – an effective introduced pathogen of the gypsy moth (*Lymantria dispar* L.) in Bulgaria. Proceedings of the International Conference “Alien Arthropods in South East Europe – Crossroad of three continents”, 19-21.09.2007, Sofia, Bulgaria, 37-43.
- PILARSKA D., V. GOLEMANSKY, D. TAKOV, M. GLAVENDEKIC, R. TOMOV 2010. Entomopathogens (Protozoans, Microsporidians and Fungi) of Alien Insects from Bulgaria: Check-list and Distribution. – *Acta zoologica bulgarica*, **62** (2): 117-130.
- PILARSKA D., A. LINDE, P. PILARSKI, G. GEORGIEV, D. TAKOV, L. SOLTER 2010. Release of *Nosema lymantriae*, *Vairimorpha disparis* and *Entomophaga maimaiga* for classical and augmentative biological control of gypsy moth in Bulgaria and the United states. – In: Proceedings of 43th Annual Meeting of the Society for Invertebrate Pathology, Trabzon, Turkey, Symposium Microsporidia “Microsporidia and other pathogens in arthropods from the Eastern Mediterranean region”, 11-15.07.2010, Trabzon, Turkey, CD, 1-6.
- PILARSKA D., R. ZIMMERMANN, A. LINDE, M. MCMANUS D. TAKOV 2001. On the occurrence of *Entomophaga auliciae* in high density browntail moth (*Euproctis chrysorrhoea* L.) populations in Bulgaria. – In: Proceedings of Third Balkan Scientific Conference, Study, Conservation and Utilisation of Forest Resources, Sofia, 2-6.10. 2001, **III**, 139-143.
- POLOVINKO G., O. YAROSLAVTSEVA, Z. TESHEBAEVA, V. KRYUKOV 2010. Dominating species of entomophilous Ascomycetes anamorphs in West Siberia, Primorsky Krai, and Kyrgyzstan. – *Contemporary problems of ecology* **3**: (5) 515-521.
- PÜNTENER W. 1981. Manual for Field Trials in Plant Protection. Second Edition. Ciba-Geigy Limited, Basle, Switzerland.
- SAMSON R. A., H. C. EVANS, J.-P. LATGE 1988. Atlas of Entomopathogenic Fungi, Springer – Verlag, London, UK, pp. 1-187.
- SEVIM A., I. DEMIR, Z. DEMIRBAG 2010. Molecular characterization and virulence of *Beauveria* spp. from the pine processionary moth, *Thaumetopoea pityocampa* (Lepidoptera: Thaumetopoeidae). – *Mycopathologia*, **170**: 269-277.
- SOLTER L., D. PILARSKA, C. VOSSBRINCK 2000. Host specificity of Microsporidia pathogenic to forest Lepidoptera. – *Biological Control*, **19** (1): 48-56.
- SOLTER L., J. SIEGEL, D. PILARSKA, C. HIGGS 2002. The impact of mixed infection of three species of microsporidia from the gypsy moth, *Lymantria dispar* L. (Lepidoptera: Lymantriidae). – *Journal of Invertebrate Pathology*, **81** (2): 103-113.
- ST. LEGER R. J., 1995. The role of cuticle-degrading proteases in fungal pathogenesis of insects. – *Canadian Journal of Botany*, **73** (Suppl.): S1119-S1125.
- ST. LEGER R. J., A. K. CHARNLEY, R. M. COOPER 1986. Cuticle-degrading enzymes of entomopathogenic fungi: Mechanisms of interaction between pathogen enzymes and insect cuticle. – *Journal of Invertebrate Pathology*, **47**: 295-302.

Received: 14.05.2011
Accepted: 12.06.2013